

Full Paper

Characterisation of 5' upstream and coding sequences of myostatin gene (*MSTN*) in Thai swamp buffalo (*Bubalus bubalis*)

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Abstract: To characterise the 5' upstream and coding sequences of the myostatin gene (*MSTN*) in Thai swamp buffalo (*Bubalus bubalis*), polymerase chain reaction and direct DNA sequencing were performed. The sequences obtained were compared with those of other livestock animals. The study successfully characterised the 5' upstream and coding sequences of the Thai swamp buffalo *MSTN* gene. Both the common and muscle-specific transcription factor binding sites were observed in the 5' upstream region and the coding sequences in the Thai swamp buffalo were similar to those in cattle (*Bos taurus*). From the multiple alignment method, the coding sequence identity of the tested buffalo was 98, 94 and 81% as compared to cattle, pig and chicken respectively. Two variants (g.264C>T and g.414G>A) located in exon 1 were also identified. Based on the obtained DNA sequences, the amino acid sequence identity of the tested buffalo was 98, 94 and 86% as compared to cattle, pig and chicken respectively. Thai swamp buffalo have 100% sequence identity of Arg-Ser-Arg-Arg (RSRR) aligned between the positions 263 and 266 of the amino acid sequence.

These results indicate that the 5' upstream and the coding sequences of Thai swamp buffalo *MSTN* gene display a high similarity with other livestock animals, especially cattle.

Keywords: myostatin gene, coding sequence, Thai swamp buffalo, *Bubalus bubalis*, transcription factor binding site

INTRODUCTION

In Thailand most buffalo (*Bubalus bubalis*) are of the swamp buffalo [1]. Traditionally, they have been raised as draught animals but more recently are increasingly being raised for meat production. At present, there is a decrease in the number of Thai swamp buffalo raised for draught purpose due to socio-economic changes, while buffalo are still used as a major meat source [2]. Buffalo meat is consumed by many Thais, especially in the north of the country, and the demand for this meat has been increasing due to an increasing population. To improve productivity, several biotechnologies have been developed, which include artificial insemination (a simple biotechnology commonly used in field work), embryo transfer and cloning.

Interestingly, there is evidence that genetic factors are associated with the muscle phenotype in livestock animals and several studies have shown that either mutation or variation in several genes is associated with meat quality [3]. The myostatin gene (*MSTN*) is an example of an economically important gene. Fluorescence *in situ* hybridisation (FISH) mapping has revealed that the buffalo *MSTN* gene is located on chromosome 2 [4]. This gene plays a critical role in regulating skeletal muscle mass. Mutations of this gene result in the double muscling phenotype in cattle [5-7]. In addition, mutation or variation identified in other livestock animals is also associated with either the growth traits or the double muscling phenotype [8-12]. However, the *MSTN* gene in swamp buffalo, particularly raised in Thailand, has not been elucidated.

The objective of this study is therefore to characterise the 5' upstream and coding sequences of Thai swamp buffalo *MSTN* gene. The characterised sequences can be used as basic data for providing molecular insights in terms of the gene's variation and mutation, including its influence on a number of biological activities.

MATERIALS AND METHODS

DNA Samples

Three millilitres each of jugular blood from eight Thai swamp buffalo kept by the Livestock Research and Breeding Centre in Surin and Lopburi provinces were collected into blood collecting tubes with ethylene diamine tetra-acetic acid as the blood coagulant [Vacuette® K3E K3EDTA, Greiner Bio-One Ltd., Thailand]. Genomic DNA was extracted from white blood cells by the salting-out method as described by Miller et al. [13]. DNA concentrations were measured using a spectrophotometer (NanoDrop 2000, Thermo Fisher Scientific, USA).

Primer Design, Polymerase Chain Reaction (PCR) and Direct DNA Sequencing

All three exons of the *MSTN* gene were amplified using overlapping primer pairs designed by primer3 software [14]. The primer design and coding sequence positions were based on DNA sequence data of the river buffalo *MSTN* gene (GenBank accession number DQ091762) [15]. Primers used for amplification of the 5' upstream sequence were designed based on cattle

MSTN gene (GenBank accession number AF348479) [16]. The regions amplified and sizes of the amplicons are shown in Table 1. The total volume of PCR reactions was 15 µL containing 50 ng of buffalo genomic DNA, 0.4 µM of each primer (Bio Basic Inc., Canada), 0.2 mM of each deoxynucleotide triphosphate (dNTPs) (Promega, UK), 4% DMSO, 2 mM MgCl₂, 1.5 µL of 10X ImmoBuffer and 0.45 unit of DNA polymerase (IMMOLASE™ DNA polymerase, Bioline USA Inc, USA). The PCR profile consisted of initial denaturation at 95°C for 10 min., 35 cycles of denaturation at 95°C for 30 sec., annealing at optimal temperature as listed in Table 1 for 30 sec. and extension at 72°C for 45 sec., and final extension at 72°C for 10 min.

The purification of PCR products was carried out using exonuclease I/shrimp alkaline phosphatase (ExoSAP-IT®, USB Corporation, USA) according to the manufacturer's instructions. Direct DNA sequencing was performed by the DNA sequencing services (First BASE DNA Sequencing Services, First BASE Laboratories, Selangor, Malaysia).

Sequence Analysis and Amino Acid Alignments

The obtained sequences from each fragment were joined together. Putative transcription binding sites in the 5' upstream region were predicted using MatInspector software [17]. Full-length coding sequences obtained from the joined fragments and generated amino acids were analysed using the Biology WorkBench 3.2 online program [18].

Table 1. List of primers used for 5' upstream and coding sequence characterisation. Fragment 1-6 primers were designed based on river buffalo *MSTN* gene (GenBank accession number DQ091762). The 5'Ups 1 and 5'Ups 2 primers were designed based on the cattle *MSTN* gene (GenBank accession number AF348479). Both GenBank accession numbers were retrieved from the NCBI database.

Name	Forward primer (5'→3')	Reverse primer (5'→3')	Product size (bp)	Annealing temperature (°C)
5'Ups 1	CTTCAAATGCTTACT TAAATAG	AATATAAACAGAGA TTTGCAGTT	575	58
5'Ups 2	AGTAATTCCATGAGC AATTTA	CTCTTGCTCCACAAT GAAT	525	58
Fragment 1 (Exon 1)	GGAAGAAGTAAGAA CAAGGGA	CTCTCTGGACATCGA ACTGA	340	60
Fragment 2 (Exon 1)	TCCTCAAGACTAGA AGCCATAA	CTCCTTACATACAAG CCAGCA	341	60
Fragment 3 (Exon 2)	GATATGGAGGTGTTC GTTCG	TGATTTCAATGCCTA AGTTGG	378	57
Fragment 4 (Exon 2)	TGAGACCTGTCAAG ACTCCT	TAAGCACAGGAAAC TGGTAG	370	60
Fragment 5 (Exon 3)	GGCATTTCAGATACTC AAACC	TGTCTGTTACCTTGA CTTCTA	293	60
Fragment 6 (Exon 3)	GGATGGGATTGGAT TATTG	AGACCTTCCATGTTT GAGG	291	60

RESULTS AND DISCUSSION

Characterisation of 5' Upstream Sequence of Buffalo *MSTN* Gene

Approximately 700 base pairs of this region were amplified using two overlapping primer pairs with annealing temperature at 58°C. By using the cattle *MSTN* gene as a reference sequence for primer designing, the 5' upstream sequence in Thai swamp buffalo is amplified and characterised (Scheme 1). Multiple alignments with those of cattle (GenBank accession number AF348479), goat (AY827576), sheep (AY918121), horse (GQ183900) and pig (AF093798) show that this region shares 99, 97, 97, 88 and 88 % sequence identity respectively. The bioinformatics analysis using MatInspector software exhibits three TATA boxes (TATA-1, TATA-2 and TATA-3). Apart from the common transcription factor binding sites, muscle-specific transcription factor binding sites (E-boxes, MEF2, SMAD and FoxO) are observed as shown in Scheme 1. In addition, we find that each transcription factor binding site shows a high conservation among the compared animal sequences, especially for ruminants. The high conservation of these transcription factor binding sites among the animals compared indicates that it may be important for transcriptional regulation of the buffalo *MSTN* gene as well as other livestock. For example, Du et al. [19] studied the transcriptional regulation activities by detecting the fluorescence strength of enhanced green fluorescent protein in C2C12 myoblasts transfected with the vectors. The results indicate that some regions such as E3 and MEF2 have an influence on the transcription and expression of the sheep *MSTN* gene. Moreover, the mutagenesis of the highly conserved FoxO or SMAD binding sites significantly decreases the *MSTN* promoter activity [20]. Actually, several transcription factor binding sites located in the 5' regulatory region of *MSTN* have been observed in other animals [21-23]. However, the 5' upstream sequence and transcription factor binding sites located in this region of both the river and swamp buffalo have not been identified before. This study is the first report describing these issues.

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-667  tatagcagaaaatccttttactagtatcacagtccttttcatttaagtcttcctgaataaat
                                     MEF2
ctgtatttttctaattatacagggaactaaaaataatttaataatacaaaaataaaattcctttt
      E-4          TATA-3
acttcaaatgcttacttaaatagttataaaatcattttattttctgagggaaaaatatatc
aactttttaagtatgaagtgtacattaagatttattcacttaaattataatttttaagt
ttcacatataaagatgaataagatctaagtgtatatgttattgttaataaagtttttaatt
ttttcgaatgtcacatacagcctttatttattcatagatttattccttttaagaagtagtc
      E-3          FoxO          SMAD
aaatgaatcagctcacccttgactgtaa caaaatactgtttggtgacttgtagcagacag
ggttttaacctctgacagcgagattcattgtggagcaagagccaatcacagatcccgacg
      E-2          TATA-2      E-1          TATA-1
acacttgctcatcaaagttggaaataaaaagccacttggaatacagataaaaagattc
actggtgtggcaagttgtctctcagactgggcaggcattaacgtttgcttggcgttact
caaaagcaaaagaaaagtaaaaggaagaagtaagaacaagggaaaagattgtattgattt
taaaaccatg +3

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Scheme 1. Results of analysis of 5' upstream sequence of Thai swamp buffalo *MSTN* gene. Putative transcription factor binding sites are indicated by boxes. The atg indicates the start codon.

Characterisation of Coding Sequences of Buffalo *MSTN* Gene

The coding sequences of the *MSTN* gene, which has been reported as a negative regulator of skeletal muscle growth [24], have previously been analysed in Brazilian Murrah buffalo [25] and Indian river buffalo [26]. In this study the characterisation of the *MSTN* coding sequences was performed in eight Thai swamp buffalo. The procedure consisted of amplification, sequencing and comparing the obtained sequences with those from cattle, pig and chicken *MSTN* genes. By using the river buffalo sequence as a reference for primer designing, the study succeeded in characterising the entire coding sequence of the *MSTN* gene (Scheme 2). The results show that the coding sequences in all 8 Thai swamp buffalo tested are identical to the river buffalo sequences (data not shown). A close relationship between the swamp and river buffalo *MSTN* genes has been previously reported. A molecular tree shows a cluster of their own and an apparent differentiation from 4 *Bos* species [27].

We also found that these sequences of the Thai buffalo *MSTN* gene are close to the *MSTN* cattle sequence rather than to the pig or chicken *MSTN* genes. In comparison to other livestock species, the entire coding sequence of Thai swamp buffalo *MSTN* gene has 98, 94 and 81% similarity to cattle, pig and chicken respectively (Scheme 3). These findings correlate well with their evolutionary relationships. Moreover, the *MSTN* gene variations are also firstly identified in Thai swamp buffalo. Two variants (g.264C>T and g.414G>A) located in exon 1 are found with the ratio of wild type to heterozygous variant samples at 5:3. The position g.264 (the number being based on river buffalo *MSTN* gene accession number DQ091762) is in codon 73 (AGC). Substitution of the third nucleotide (AGC>AGT) does not result in any change of the amino acid (serine). Likewise, the G>A transition at genomic DNA position g.414 (ACG>ACA) does not result in an alteration of threonine at codon position 123.

In their previous studies of river buffalo, Mota et al. [25] and Tantia et al. [26] did not find any variant in coding regions among the buffalo. It can thus be suggested that both the identified variants in our study may be specific for Thai swamp buffalo. We suggest that, as DNA variations in genomic DNA positions 264 and 414 are identified with high frequencies, these can be used as molecular markers for buffalo identification.

Based on the nucleic acid sequences obtained for the buffalo *MSTN* gene, amino acids were generated using the Biology WorkBench 3.2. It was found that the amino acid sequence of Thai swamp buffalo is 98, 94 and 86% identical to that of cattle, pig and chicken respectively (Scheme 4). To generate a biologically active myostatin ligand (a COOH-terminal fragment), cleavage by the furin-family enzyme at Arg-Ser-Arg-Arg (RSRR) aligned between positions 263 and 266 of amino acids is needed [31]. Although DNA sequences encoding for RSRR are not fully conserved among the species shown in Scheme 3, these sequences encode the same amino acid sequence. Analysis results in this study have revealed that this four-amino-acid region of Thai swamp buffalo is 100% identical to that of cattle, pig and chicken. In addition, a COOH-terminal fragment of Thai swamp buffalo shows 99, 97 and 96% amino acid similarity to myostatin of cattle, pig and chicken respectively. These results indicate that the myostatin of Thai swamp buffalo may have the same function as in other livestock animals. In this study although we did not find missense, nonsense and frameshift mutations (which can result in increased muscle mass) in any of the Thai swamp buffalo tested, further study on a larger number of animals should be done to clarify the possibility of mutation.

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      M Q K L Q I S V Y I Y L F M L I V A G P
1  atgcaaaaactgcaaattctctgtttatattttacctatgtgattgttgctggccca 60
    V D L N E N S E Q K E N V E K E G L C N
61  gtggatctgaatgagaacagcagcagaaggaatgtggaaaaagaggggtgtgtaat 120
    A C L W R E N T T S S R L E A I K I Q I
121  gcatgtttgtggagggaaaaactacatcctcaagactagaagccataaaaaatccaaatc 180
    L S K L R L E T A P N I S K D A I R Q L
181  ctgagtaaaacttcgcctggaaacagctcctaacatcagcaaagatgctatcagacaactt 240
    L P K A P P L L E L I D Q F D V Q R D A
241  ttgcccaaggctcctccactcctggaaactgattgatcagttcgatgtccagagagatgcc 300
    G S D G S L E D D D Y H A R T D A V I T
301  ggcagtgcaggctccttgggaagacgatgactaccacgccaggacggacgcgggtcattacc 360
    M P T E S D L L T Q V E G K P K C C F F
361  atgccacggagtctgatcttctaacgcaagtgggaaggaaaacccaaatgttgcttcttt 420
    Q F S S K I Q Y N K L V K A Q L W I L
421  caatttagctctaagatacaatacaataaactgtaaaggcccaactgtggatatatctg 480
    R P V K T P A T V F V Q I L R L I K P M
481  agaccctgcaagactcctgcgacagtgtttgtgcagatcctgagactcatcaaaccatg 540
    K D G T R Y T G I R S L K L D M N P G T
541  aaagacgggtacaaggtatactggaatccgatctctgaaacttgacatgaaccaggcact 600
    G I W Q S I D V K T V L Q N W L K Q P E
601  ggtatttggcagagcattgatgtgaagacagtgttgcaaaactggctcaaacaacctgaa 660
    S N L G I E I K A L D E N G H D L A V T
661  tccaacttaggcattgaaatcaaagcttttagatgagaatggatcatgattgtgtaacc 720
    F P E P G E D G L T P F L E V K V T D T
721  ttcccagaaccaggagaagatggactgactccttttttagaagtcaaggtaacagacaca 780
    P K R S R R D F G L D C D E R S T E S R
781  ccaaaaagatctaggagagattttgggcttgattgtgatgagcgctccacagaatctcga 840
    C C R Y P L T V D F E A F G W D W I I A
841  tgctgtcggttacccctctaactgtggattttgaagcttttggatgggattggattattgca 900
    P K R Y K A N Y C S G E C E F V F L Q K
901  cctaaaagatataaggccaattactgctctggagaatgtgaatttgtatttttgcaaaag 960
    Y P H T H L V H Q A N P R G S A G P C C
961  tatcctcataccatcttgtgcaccaagcaaaccacagaggttcagctggcccctgctgc 1020
    T P T K M S P I N M L Y F N G E G Q I I
1021  actcctacaaagatgtctccaattaatatgctatattttaatggcgaaggacaaataata 1080
    Y G K I P A M V V D R C G C S *
1081  tatgggaagattccagccatggtagtagatcgctgtgggtgttcatga 1128

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Scheme 2. Coding and amino acid sequences of Thai swamp buffalo *MSTN* gene. The first codon is underlined. Asterisk indicates stop codon.

Swamp buffalo	1	<u>ATGCAAAA</u> ACTGCAAATCTCTGCTTATATTACCTATTTATGCTGATTGTTGCTGGCCCA
Cattle		ATGCAAAA
Pig		ATGCAAAA
Chicken		ATGCAAAAGCTAGCAGTCTATGTTTATATTACCTGTTTATGCTGAGATCGCGGTTGATCCA
		***** * * * * *
Swamp buffalo	61	GTGGATCTGAATGAGAACAGCAGCAGCAGAAAGGAAAATGTGAAAAAGAGGGGCTGTGTAAT
Cattle		GTGGATCTGAATGAGAACAGCAGCAGCAGAAAGGAAAATGTGAAAAAGAGGGGCTGTGTAAT
Pig		GTGGATCTGAATGAGAACAGCAGCAGCAGAAAGGAAAATGTGAAAAAGAGGGGCTGTGTAAT
Chicken		GTGGCTCTGGATGCGAGTAGTCAGCCACAGAGAACGCTGAAAAAGACGGACTGTGCAAT
		**** * * * * *
Swamp buffalo	121	GCATGTTTGTGGAGGGAAAAGCACTACATCCTCAAGACTAGAAGCCATAAAAAATCCAAATC
Cattle		GCATGTTTGTGGAGGGAAAAGCACTACATCCTCAAGACTAGAAGCCATAAAAAATCCAAATC
Pig		GCATGTATGTGGAGACAAAACACTAAATCTTCAAGACTAGAAGCCATAAAAAATCCAAATC
Chicken		GCTTGTACGTGGAGACAGAATACAAAATCCTCCAGAAATAGAAGCCATAAAAAATCCAAATC
		** * * * * * * * * * *
Swamp buffalo	181	CTCAGTAAACTTCGCCTGGAAACAGCTCCTAACATCAGCAAAGATGCTATCAGACAACCTT
Cattle		CTCAGTAAACTTCGCCTGGAAACAGCTCCTAACATCAGCAAAGATGCTATCAGACAACCTT
Pig		CTCAGTAAACTTCGCCTGGAAACAGCTCCTAACATTAGCAAAGATGCTATAAGACAACCTT
Chicken		CTCAGCAAACCTGCCTGGAAACAGCACCTAACATTAGCAGGGACGTTATTAAGCAGCTT
		***** * * * * *
Swamp buffalo	241	TTGCCCAAGGCTCCTCCACTCCTGGAAGTGAATGATCAGTTCGATGTCCAGAGAGATGCC
Cattle		TTGCCCAAGGCTCCTCCACTCCTGGAAGTGAATGATCAGTTCGATGTCCAGAGAGATGCC
Pig		TTGCCCAAGGCTCCTCCACTCGGGAACTGATGATCAGTACGATGTCCAGAGAGATGAC
Chicken		TTACCCAAAGCTCCTCCACTGCAGGAAGTGAATGATCAGTATGATGTCCAGAGGGACGAC
		** * * * * * * * * * *
Swamp buffalo	301	GGCAGTGACGGCTCCTTGGAAAGCAGTGAATACACGCCAGGACGGACGCGGTGATTACC
Cattle		AGCAGTGACGGCTCCTTGGAAAGCAGTGAATACACGCCAGGACGGAAACGGTATTACC
Pig		AGCAGTGATGGCTCCTTGGAAAGATGATGATTATCAGCTACGACGGAAACGATATTACC
Chicken		AGTAGCGATGGCTCTTTGGAAGACGATGACTATCATGCCACAACCGAGACGATTATCACA
		* * * * * * * * * * *
Swamp buffalo	361	ATGCCACGGAGTCTGATCTTCTAACGCAAGTGGAAAGGAAAAACCAATGTTGCTTCTTT
Cattle		ATGCCACGGAGTCTGATCTTCTAACGCAAGTGGAAAGGAAAAACCAATGTTGTTTCTTT
Pig		ATGCCTACAGAGTCTGATCTTCTAATGCAAGTGGAAAGGAAAAACCAATGCTGCTTCTTT
Chicken		ATGCCTACGGAGTCTGATTTTCTTGACAAATGGAGGGAAAAACCAATGTTGCTTCTTT
		***** ** * * * * *
Swamp buffalo	421	CAATTTAGCTCTAAGATACAATACAATAAACTAGTAAAGGCCCAACTGTGGATATATCTG
Cattle		AAATTTAGCTCTAAGATACAATACAATAAACTAGTAAAGGCCCAACTGTGGATATATCTG
Pig		AAATTTAGCTCTAATAACAATAACAATAAAGTAGTAAAGGCCCAACTGTGGATATATCTG
Chicken		AAGTTTAGCTCTAATAACAATAACAATAAAGTAGTAAAGGCACAATTATGGATATACTTG
		* * * * * * * * * * *
Swamp buffalo	481	AGACCCGTCAAGACTCCTGCGACAGTGTGTTGTGCAGATCCTGAGACTCATCAAACCCATG
Cattle		AGGCCGTCAAGACTCCTGCGACAGTGTGTTGTGCAATCCTGAGACTCATCAAACCCATG
Pig		AGACCCGTCAAGACTCCTACAACAGTGTGTTGTGCAATCCTGAGACTCATCAAACCCATG
Chicken		AGGCAAGTCAAAAACCTACAACGGTGTGTTGTGCAGATCCTGAGACTCATTAAGCCCATG
		** * * * * * * * * * *
Swamp buffalo	541	AAAGACGGTACAAGGTATACTGGAATCCGATCTCTGAAACTTGACATGAACCCAGGCACT
Cattle		AAAGACGGTACAAGGTATACTGGAATCCGATCTCTGAAACTTGACATGAACCCAGGCACT
Pig		AAAGACGGTACAAGGTATACTGGAATCCGATCTCTGAAACTTGACATGAACCCAGGCACT
Chicken		AAAGACGGTACAAGGTATACTGGAATCCGATCTCTGAAACTTGACATGAACCCAGGCACT
		***** * * * * *

Scheme 3. Multiple alignments of *MSTN* coding sequences in four livestock animals: swamp buffalo, cattle (GenBank accession number AY160688) [28], pig (AY448008) [29] and chicken (AY448007) [30]. The first codon is underlined. The nucleotide positions 1126-1128 are stop codon. Asterisks indicate consensus sequence between four species. Box indicates variations of DNA sequence among species encoding amino acids for cleavage site (RSRR).

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Swamp buffalo 601 GGTATTTGGCAGAGCATTGATGTGAAGACAGTGTTGCAAAACTGGCTCAAAACAACCTGAA
Cattle          GGTATTTGGCAGAGCATTGATGTGAAGACAGTGTTGCGAAGCTGGCTCAAAACAACCTGAA
Pig            GGTATTTGGCAGAGCATTGATGTGAAGACAGTGTTGCAAAATTGGCTCAAAACAACCTGAA
Chicken        GGTATCTGGCAGAGTATTGATGTGAAGACAGTGCTGCAAAATTGGCTCAAAACAGCCTGAA
                *****

Swamp buffalo 661 TCCAACCTTAGGCATTGAAATCAAAGCTTTAGATGAGAATGGTCATGATCTTGCTGTAACC
Cattle          TCCAACCTTAGGCATTGAAATCAAAGCTTTAGATGAGAATGGCCATGATCTTGCTGTAACC
Pig            TCCAACCTTAGGCATTGAAATCAAAGCTTTAGATGAGAATGGTCATGATCTTGCTGTAACC
Chicken        TCCAATTTAGGCATCGAAATAAAAGCTTTTGATGAGACTGGACGAGATCTTGCTGTCACA
                *****

Swamp buffalo 721 TTCCCAGAACCCAGGAGAAGATGGACTGACTCCTTTTGAAGTCAAGGTAACAGACACA
Cattle          TTCCCAGAACCCAGGAGAAGATGGACTGACTCCTTTTGAAGTCAAGGTAACAGACACA
Pig            TTCCCAGAACCCAGGAGAAGATGGGCTGAATCCCTTTTGAAGTCAAGGTAACAGACACA
Chicken        TTCCCAGAACCCGGGTGAAGATGGATTGAACCCATTTTGAAGGTCAGAGTTACAGACACA
                *****

Swamp buffalo 781 CCAAAAGATCTAGGAGAGATTTGGGCTTGATTGTGATGAGCGCTCCACAGAATCTCGA
Cattle          CCAAAAGATCTAGGAGAGATTTGGGCTTGATTGTGATGAACACTCCACAGAATCTCGA
Pig            CCAAAAGATCTAGGAGAGATTTGGGCTTGATTGTGATGAGCACTCAACAGAATCTCGA
Chicken        CCGAAACGGTCCCGCAGAGATTTGGGCTTGATTGTGATGAGCACTCAACGGAATCCCGA
                ****

Swamp buffalo 841 TGCTGTCGTTACCCCTCTAAGCTGTGGATTTTGAAGCTTTTGGATGGGATTGGATTATTGCA
Cattle          TGCTGTCGTTACCCCTCTAAGCTGTGGATTTTGAAGCTTTTGGATGGGATTGGATTATTGCA
Pig            TGCTGTCGTTACCCCTCTAAGCTGTGGATTTTGAAGCTTTTGGATGGGATTGGATTATTGCA
Chicken        TGTTGTCGCTACCCGCTGACAGTGGATTTCGAAGCTTTTGGATGGGATTGGATTATTGCA
                *****

Swamp buffalo 901 CCTAAAAGATATAAGGCCAATTACTGCTCTGGAGAATGTGAATTTGTATTTTGCAAAAG
Cattle          CCTAAAAGATATAAGGCCAATTACTGCTCTGGAGAATGTGAATTTGTATTTTGCAAAAG
Pig            CCTAAAAGATATAAGGCCAATTACTGCTCTGGAGAATGTGAATTTGTATTTTGCAAAAG
Chicken        CCTAAAAGATATAAGGCCAATTACTGCTCTGGAGAATGTGAATTTGTATTTTGCAAAAG
                *****

Swamp buffalo 961 TATCCTCATACCCATCTTGTGCACCAAGCAAACCCAGAGGTTTACGCTGGCCCTGCTGC
Cattle          TATCCTCATACCCATCTTGTGCACCAAGCAAACCCAGAGGTTTACGCTGGCCCTGCTGC
Pig            TATCCTCATACCCATCTTGTGCACCAAGCAAACCCAGAGGTTTACGCTGGCCCTGCTGC
Chicken        TACCCGCACACTCACCTGGTACACCAAGCAAATCCAGAGGCTCAGCAGGCCCTTGTGCTGC
                *****

Swamp buffalo 1021 ACTCCTACAAAGATGTCTCCAATTAATATGCTATATTTTAAATGGCGAAGGACAAATAATA
Cattle          ACTCCTACAAAGATGTCTCCAATTAATATGCTATATTTTAAATGGCGAAGGACAAATAATA
Pig            ACTCCTACAAAGATGTCTCCAATTAATATGCTATATTTTAAATGGCGAAGGACAAATAATA
Chicken        ACACCCACCAAGATGTCCCTATAAACATGCTGTATTTCAATGGAAGAAACAAATAATA
                *****

Swamp buffalo 1081 TATGGGAAGATTCCAGCCATGGTAGTAGATCGCTGTGGGTGTTTCATGA 1128
Cattle          TATGGGAAGATTCCAGCCATGGTAGTAGATCGCTGTGGGTGTTTCATGA
Pig            TATGGGAAGATTCCAGCCATGGTAGTAGATCGCTGTGGGTGTTTCATGA
Chicken        TATGGGAAGATACCAGCCATGGTAGTAGATCGCTGTGGGTGTTTCATGA
                *****

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Scheme 3 (Continued).

Swamp buffalo	1	MQKLQISVYIYLFMLIVAGPVDLNENSEQKENVEKEGLCNACLWRENTTSSRLEAIKIQI	60
Cattle		MQKLQISVYIYLFMLIVAGPVDLNENSEQKENVEKEGLCNACLWRENTTSSRLEAIKIQI	
Pig		MQKLQIYVYIYLFMLIVAGPVDLNENSEQKENVEKEGLCNACMWRQNTKSSRLEAIKIQI	
Chicken		MQKLAVYVYIYLFMQIAVDFVALDGSSQPTENAEKDGLCNACTWRQNTKSSRIEAIKIQI	
		**** * * * * *	
Swamp buffalo	61	LSKLRLLETAPNISKDAIRQLLPKAPPLLELIDQFDVQRDAGSDGSLEDDDYHARTDAVIT	120
Cattle		LSKLRLLETAPNISKDAIRQLLPKAPPLLELIDQFDVQRDASSDGSLEDDDYHARTETVIT	
Pig		LSKLRLLETAPNISKDAIRQLLPKAPPLRELIDQYDVQRDDSSDGSLEDDDYHATTETIIT	
Chicken		LSKLRLLEQAPNISRDVIKQLLPKAPPLQELIDQYDVQRDDSSDGSLEDDDYHATTETIIT	
		***** * * *	
Swamp buffalo	121	MPTESDLLTQVEGKPKCCFFQFSSKIQYNKLVKAQLWIYLRPVKTPATVFVQILRLIKPM	180
Cattle		MPTESDLLTQVEGKPKCCFFQFSSKIQYNKLVKAQLWIYLRPVKTPATVFVQILRLIKPM	
Pig		MPTESDLLMQVEGKPKCCFFQFSSKIQYNKLVKAQLWIYLRPVKTPPTTFVQILRLIKPM	
Chicken		MPTESDFLVQMEGKPKCCFFQFSSKIQYNKLVKAQLWIYLRQVQKPTTFVQILRLIKPM	
		***** * * *	
Swamp buffalo	181	KDGTRYTGIRSLKLDMPGTGIWQSIDVKTVLQNLKQPESNLGIEIKALDENGHD LAVT	240
Cattle		KDGTRYTGIRSLKLDMPGTGIWQSIDVKTVLQNLKQPESNLGIEIKALDENGHD LAVT	
Pig		KDGTRYTGIRSLKLDMPGTGIWQSIDVKTVLQNLKQPESNLGIEIKALDENGHD LAVT	
Chicken		KDGTRYTGIRSLKLDMPGTGIWQSIDVKTVLQNLKQPESNLGIEIKAFDETGRDLAVT	
		***** * * *	
Swamp buffalo	241	FPEPGEDGLTPFLEVKTDTPKRSRRDFGLDCDERSTESRCCRYPLTVDFEAFGWDWIIA	300
Cattle		FPEPGEDGLTPFLEVKTDTPKRSRRDFGLDCDERSTESRCCRYPLTVDFEAFGWDWIIA	
Pig		FPGPGEDGLNPFLEVKTDTPKRSRRDFGLDCDERSTESRCCRYPLTVDFEAFGWDWIIA	
Chicken		FPGPGEDGLNPFLEVRVDTPKRSRRDFGLDCDERSTESRCCRYPLTVDFEAFGWDWIIA	
		** ***** *	
Swamp buffalo	301	PKRYKANYCSGECEFVFLQYPHTLVHQANPRGSAGPCCTPTKMSPINMLYFNGEQII	360
Cattle		PKRYKANYCSGECEFVFLQYPHTLVHQANPRGSAGPCCTPTKMSPINMLYFNGEQII	
Pig		PKRYKANYCSGECEFVFLQYPHTLVHQANPRGSAGPCCTPTKMSPINMLYFNGEQII	
Chicken		PKRYKANYCSGECEFVFLQYPHTLVHQANPRGPAGPCCTPTKMSPINMLYFNGEQII	
		***** * * *	
Swamp buffalo	361	YGKIPAMVVDRCGCS	375
Cattle		YGKIPAMVVDRCGCS	
Pig		YGKIPAMVVDRCGCS	
Chicken		YGKIPAMVVDRCGCS	

Scheme 4. Multiple alignment of myostatin amino acid sequences in four livestock animals: swamp buffalo, cattle (GenBank accession number AF320998) [32], pig (EF490986) [33] and chicken (AF346599) [34]. The cleavage site (RSRR) is shown in the box. Asterisks indicate consensus amino acid residues between the four species.

CONCLUSIONS

Our results have demonstrated that the 5' upstream, coding and amino acid sequences of the Thai swamp buffalo *MSTN* gene show high sequence identity with other livestock animals, especially cattle. To our knowledge, this is the first *MSTN* gene report for Thai swamp buffalo. The output provides basic information that can be used in further study such as buffalo breeding.

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REFERENCES

1. K. Triwitayakorn, B. Moolmuang, S. Sraphet, S. Panyim, A. Na-Chiangmai and D. R. Smith, "Analysis of genetic diversity of the Thai swamp buffalo (*Bubalus bubalis*) using cattle microsatellite DNA markers", *Asian-Australas. J. Anim. Sci.*, **2006**, 19, 617-621.
2. S. Uriyapongson, "Buffalo and buffalo meat in Thailand", *Buffalo Bull.*, **2013**, 32, 329-332.
3. H. K. Koopaei and A. E. Koshkoiyeh, "Application of genomic technologies to the improvement of meat quality in farm animals", *Biotechnol. Mol. Biol. Rev.*, **2011**, 6, 126-132.
4. M. Strazzullo, C. Rossetti, G. Fusco, C. Campanile, D. Vecchio, G. Campanile, A. Perucatti, G. P. Di Meo, F. Filippini, A. Eggen, L. Ferrara and M. D'Esposito, "Genomic characterization and chromosomal mapping of 5 river buffalo skeletal muscle differentiation master genes", *Cytogenet. Genome Res.*, **2010**, 128, 221-227.
5. C. Marchitelli, M. C. Savarese, A. Crisa, A. Nardone, P. A. Marsan and A. Valentini, "Double muscling in Marchigiana beef breed is caused by a stop codon in the third exon of myostatin gene", *Mamm. Genome.*, **2003**, 14, 392-395.
6. J. L. Gill, S. C. Bishop, C. McCorquodale, J. L. Williams and P. Wiener, "Associations between the 11-bp deletion in the myostatin gene and carcass quality in Angus-sired cattle", *Anim. Genet.*, **2009**, 40, 97-100.
7. P. Wiener, J. A. Woolliams, A. Frank-Lawale, M. Ryan, R. I. Richardson, G. R. Nute, J. D. Wood, D. Homer and J. L. Williams, "The effects of a mutation in the myostatin gene on meat and carcass quality", *Meat Sci.*, **2009**, 83, 127-134.
8. I. A. Boman, G. Klemetsdal, T. Blichfeldt, O. Nafstad and D. I. Vage, "A frameshift mutation in the coding region of the myostatin gene (*MSTN*) affects carcass conformation and fatness in Norwegian White Sheep (*Ovis aries*)", *Anim. Genet.*, **2009**, 40, 418-422.
9. I. A. Boman, G. Klemetsdal, O. Nafstad, T. Blichfeldt and D. I. Vage, "Impact of two myostatin (*MSTN*) mutations on weight gain and lamb carcass classification in Norwegian White Sheep (*Ovis aries*)", *Genet. Sel. Evol.*, **2010**, 42, doi:10.1186/1297-9686-42-4
10. T. Tozaki, F. Sato, E. W. Hill, T. Miyake, Y. Endo, H. Kakoi, H. Gawahara, K. Hirota, Y. Nakano, Y. Nambo and M. Kurosawa, "Sequence variants at the myostatin gene locus influence the body composition of Thoroughbred horses", *J. Vet. Med. Sci.*, **2011**, 73, 1617-1624.
11. G. Zhang, L. Zhang, Y. Wei, J. Wang, F. Ding, G. Dai and K. Xie, "Polymorphisms of the myostatin gene and its relationship with reproduction traits in the Bian chicken", *Anim. Biotechnol.*, **2012**, 23, 184-193.
12. Z. J. Zhang, Y. H. Ling, L. J. Wang, Y. F. Hang, X. F. Guo, Y. H. Zhang, J. P. Ding and X. R. Zhang, "Polymorphisms of the myostatin gene (*MSTN*) and its relationship with growth traits in goat breeds", *Genet. Mol. Res.*, **2013**, 12, 965-971.
13. S. A. Miller, D. D. Dykes and H. F. Polesky, "A simple salting out procedure for extracting DNA from human nucleated cells", *Nucleic Acids Res.*, **1988**, 16, 1215.

14. T. Koressaar and M. Remm, "Enhancement and modifications of primer design program primer3", *Bioinformatics*, **2007**, 23, 1289-1291.
15. National Centre of Biotechnology Information (NCBI), "*Bubalus bubalis* myostatin gene, complete cds", <http://www.ncbi.nlm.nih.gov/nuccore/DQ091762> (Accessed: January 2014).
16. National Centre of Biotechnology Information (NCBI), "*Bos taurus* myostatin gene", <http://www.ncbi.nlm.nih.gov/nuccore/AF348479> (Accessed: January 2014).
17. Genomatix, "Genomatix software suite", <http://www.genomatix.de>, (Accessed: January 2014).
18. San Diego Supercomputer Centre (SDSC), "Biology WorkBench version 3.2", <http://workbench.sdsc.edu>, (Accessed: January 2014).
19. R. Du, X. R. An, Y. F. Chen and J. Qin, "Some motifs were important for myostatin transcriptional regulation in sheep (*Ovis aries*)", *J. Biochem. Mol. Biol.*, **2007**, 40, 547-553.
20. D. L. Allen and T. G. Unterman, "Regulation of myostatin expression and myoblast differentiation by FoxO and SMAD transcription factors", *Am. J. Physiol. Cell Physiol.*, **2007**, 292, 188-199.
21. D. L. Allen and M. Du, "Comparative functional analysis of the cow and mouse myostatin genes reveals novel regulatory elements in their upstream promoter regions", *Comp. Biochem. Physiol. B Biochem. Mol. Biol.*, **2008**, 150, 432-439.
22. S. Dall'Olio, L. Fontanesi, L. N. Costa, M. Tassinari, L. Minieri and A. Falaschini, "Analysis of horse myostatin gene and identification of single nucleotide polymorphisms in breeds of different morphological types", *J. Biomed. Biotechnol.*, **2010**, 2010, Art. ID 542945.
23. J. Li, J. Deng, S. Yu, J. Zhang, D. Cheng and H. Wang, "The virtual element in proximal promoter of porcine myostatin is regulated by myocyte enhancer factor 2C", *Biochem. Biophys. Res. Commun.*, **2012**, 419, 175-181.
24. A. C. McPherron and S. J. Lee, "Double muscling in cattle due to mutations in the myostatin gene", *Proc. Natl. Acad. Sci. USA.*, **1997**, 94, 12457-12461.
25. L. S. L. S. Mota, R. A. Curi, D. A. Palmieri, A. S. Borges, C. R. Lopes, J. D. Babosa and M. A. Gemines, "Sequence characterization of coding regions of the myostatin gene (*GDF8*) from Brazilian Murrah buffaloes (*Bubalus bubalis*) and comparison with the *Bos taurus* sequence", *Genet. Mol. Biol.*, **2006**, 29, 79-82.
26. M. S. Tania, R. K. Vijn, B. Mishra and S. T. B. Kumar, "Sequence of *GDF8* (Myostatin) gene in *Bubalus bubalis*", *Anim. Biotechnol.*, **2007**, 18, 177-181.
27. L. P. Wang, R. Q. Geng, D. J. Ji, H. Chang, C. F. Chang and Y. H. Li, "Molecular evolution character of complete coding sequence of bovine myostatin gene", *Acta Agric. Bor. Sin.*, **2009**, 24, 46-49.
28. National Centre of Biotechnology Information (NCBI), "*Bos taurus* myostatin (*GDF8*) mRNA, complete cds", <http://www.ncbi.nlm.nih.gov/nuccore/AY160688> (Accessed: January 2014).
29. National Centre of Biotechnology Information (NCBI), "*Sus scrofa* myostatin (*MSTN*) mRNA, complete cds", <http://www.ncbi.nlm.nih.gov/nuccore/AY448008> (Accessed: January 2014).
30. National Centre of Biotechnology Information (NCBI), "*Gallus gallus* myostatin (*MSTN*) mRNA, complete cds", <http://www.ncbi.nlm.nih.gov/nuccore/AY448007> (Accessed: January 2014).
31. Z. Huang, X. Chen and D. Chen, "Myostatin: A novel insight into its role in metabolism, signal pathways, and expression regulation", *Cell. Signal.*, **2011**, 23, 1441-1446.
32. National Centre of Biotechnology Information (NCBI), "*Bos taurus* myostatin (*GDF8*) gene, complete cds", <http://www.ncbi.nlm.nih.gov/nuccore/AF320998> (Accessed: January 2014).

33. National Centre of Biotechnology Information (NCBI), “*Sus scrofa* breed Large White myostatin (*MSTN*) gene, complete cds”, <http://www.ncbi.nlm.nih.gov/nuccore/EF490986> (Accessed: January 2014).
34. National Centre of Biotechnology Information (NCBI), “*Gallus gallus* myostatin (*MSTN*) gene, promoter region, exons 1, 2 and 3 and complete cds”, <http://www.ncbi.nlm.nih.gov/nuccore/AF346599> (Accessed: January 2014).